

RESEARCH ARTICLE

Open Access



LncRNA MIR17HG inhibits non-small cell lung cancer by upregulating miR-142-3p to downregulate Bach-1

Sen Wei^{*†}, Jinghao Liu[†], Xin Li and Xingyu Liu

Abstract

Background: This study aimed to investigate the role of MIR17HG in non-small cell lung cancer (NSCLC).

Methods: Differential expression of MIR17HG in NSCLC was first detected by exploring the TCGA dataset. Expression levels of miR-142-3p in both NSCLC and non-tumor tissues were determined by qPCR. The effects of overexpressing MIR17HG on the methylation of miR-142 were assessed by MSP. The effects of overexpressing MIR17HG, miR-142-3p and Bach-1 on the invasion and migration of NSCLC cells were assessed by Transwell invasion or migration assay.

Results: Analysis of TCGA dataset revealed slightly downregulated expression of MIR17HG in NSCLC. This downregulation was further confirmed by measuring the expression levels of MIR17HG in NSCLC and non-tumor tissues from NSCLC patients. MIR17HG was found to decrease the methylation of miR-142-3p, and overexpression of MIR17HG led to upregulated miR-142-3p. Moreover, overexpression of MIR17HG also led to downregulated Bach-1, the downstream target of miR-142-3p. Cell invasion and migration analysis showed that overexpression of MIR17HG and miR-142-3p led to inhibited cancer cell invasion and migration. In contrast, overexpression of Bach-1 played an opposite role and attenuated the effects of overexpressing MIR17HG and miR-142-3p.

Conclusion: MIR17HG inhibits NSCLC by upregulating miR-142-3p to downregulate Bach-1.

Trial registration: TJ-MU-2012-0148594, registered January 2, 2012

Keywords: Non-small cell lung cancer, MIR17HG, miR-142-3p, Bach-1

Background

Lung cancer has been the most common type of malignancy that causes the highest mortality rate among all types of malignancies for decades [1]. In 2018, lung cancer affected 11.6% (2,093,876 cases) of all newly diagnosed cancer cases and caused 18.4% (1,761,007 cases) of all cancer deaths [2]. Avoidance of smoking and second-hand smoking can significantly reduce the risk of lung cancer [3]. However, to quit smoking, it requires intensive intervention for smokers' life. More importantly, lung cancer also affects never-smokers

[4]. The current survival of lung cancer patients is still poor, largely due to the lack of effective therapies for advanced lung cancer and the low rate of early diagnosis [5, 6].

Genetic factors play central roles in the pathogenesis of lung cancer [7]. The identification of genetic factors involved in this disease provided potential targets for the development of targeted therapies [8, 9]. Non-coding RNAs (ncRNAs) have no protein-coding capacity but affect protein synthesis by regulating gene expression [10]. For instance, long (> 200 nt) non-coding RNAs (lncRNAs) are usually spatially expressed and affect local gene expression [11]. However, the functions of most lncRNAs in cancer remain unclear. LncRNA miR-17-92a-1 cluster host gene (MIR17HG) has been reported

* Correspondence: senwei03@aliyun.com

[†]Sen Wei and Jinghao Liu contributed equally to this work.

Department of Lung Cancer Surgery, General Hospital of Tianjin Medical University, No. 154 An'shan Road, Tianjin 300052, China



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

to produce cancer-related miRNAs [12]. For instance, MIR17HG produces miRNA-19a/b to mediate the anti-neoplastic effects induced by grape seed procyanidin extract in the treatment of lung cancer [13]. MIR17HG was also reported to promote colorectal cancer by interacting with miR-17-5p [14]. In the present study, we observed downregulated expression pattern of MIR17HG in NSCLC by analyzing TCGA dataset. In addition, our preliminary deep sequencing data revealed positive correlation between the expression of MIR17HG and miR-142-3p, a tumor suppressive miRNA [15], across NSCLC tissue samples (data not shown). This study aimed to investigate the potential involvement of MIR17HG in non-small cell lung cancer (NSCLC), a major subtype of lung cancer, and explore its possible interactions with miR-142-3p.

Materials and methods

Patients and tissue samples

Paired non-tumor and NSCLC tissue samples were obtained from 60 NSCLC patients (41 males and 19 females, 42 to 68 years old, mean age 54.3 ± 5.8 years old). These patients were admitted by General Hospital of Tianjin Medical University between March 2012 and May 2014. This study passed the review board of the Ethics Committee of aforementioned hospital. All patients were diagnosed for the first time. No therapies for any clinical disorders were performed within 3 months before admission. Patients complicated with other clinical disorders were excluded from this study. Based on AJCC clinical staging system, the 60 patients included 14, 16 and 30 at clinical stage II, III, and IV, respectively. All patients were informed of the experimental design of this project and signed the informed consent.

NSCLC cells and transfection

This study used human NSCLC cell lines H2170 and H2126 as the cell model of NSCLC. No mutations were observed in these two cell lines. Cells were obtained from ATCC (USA). Cells were cultured in cell culture medium composed of 90% RPMI-1640 medium and 10% FBS at 37 °C with 95% humidity and 5% CO₂. Cells were harvested at confluence of 70–80% for transfections. MIR17HG and Bach-1 expression vectors were constructed using pcDNA3.1 vector as the backbone. Negative control (NC) miRNA and miR-142-3p mimic were synthesized by RIBOBIO (Guangzhou, China). Lipofectamine 2000 (Invitrogen, USA) was used to transfect 10 nM vector (NC group was empty pcDNA3.1 vector transfection) or 45 nM miRNA (NC group was NC miRNA transfection). Untransfected cells were used as the control cells (C). The following experiments were conducted 24 h post-transfections.

RNA extractions and qRT-PCR

Ribozol (Sigma-Aldrich) was used to perform all RNA extractions following the manufacture's instructions. RNA samples were precipitated using 85% ethanol to harvest miRNAs. All RNA samples were digested by DNA eraser (Takara, USA). SSRT IV system (Thermo Fisher Scientific) was used to transcribe total RNAs into cDNAs. With cDNA samples as template, SYBR® Green Realtime PCR Master Mix (Toyobo, Japan) was used to prepare all qPCR mixtures. GAPDH was used as the endogenous control to measure the expression levels of MIR17HG and Bach-1. All-in-One™ miRNA qRT-PCR Detection Kit (GeneCopoeia) was used to measure the expression levels of mature miR-142-3p. All steps including polyadenylation, reverse transcriptions and qPCR assays were performed following the instructions from GeneCopoeia. U6 was used as the endogenous control. All qPCR assays were repeated 3 times and data normalizations were performed using $2^{-\Delta\Delta C_q}$ method.

Methylation specific PCR (MSP)

Genomic DNAs were extracted from H2170 cells at 24 h post-transfection using the Wizard® Genomic DNA Purification Kit (Promega Corporation). EZ DNA Methylation-Gold™ Kit (ZYMO RESEARCH, USA) was used to convert DNA samples. DNA methylation was measured by performing PCR reactions using the Taq 2X Master Mix (NEB).

Transwell assays

The effects of transfections on the invasion and migration of cells were evaluated by Transwell invasion and migration assays, respectively. Transwell insert (Corning) was used for cell migration assay. The upper chamber was coated with Matrigel for cell invasion assay. The upper chamber was filled with 500 µl non-serum cell suspension containing 10⁵ cells, while the lower chamber was filled with a mixture of 20% FBS and 80% aforementioned medium. Cells were cultivated under aforementioned conditions for 12 h, followed by staining of the lower membrane surface using 0.5% crystal violet (Sigma-Aldrich). The stained cells were observed under a light microscope.

Western blot

RIPA solution (Sigma-Aldrich) was used for protein extractions. All proteins samples were incubated in boiling water for 10 min for denaturation. Electrophoresis was then performed using 12% SDS-PAGE gel. After that, protein samples were transferred to PVDF membranes, followed by blocking in PBS containing 5% non-fat milk. The blocked membranes were then incubated with primary antibodies of rabbit anti-GAPDH (ab9485, 1:1400, Abcam) and

Bach-1 (ab7288, 1:1400, Abcam) primary antibodies. IgG-HRP secondary antibody (ab6721, 1:1400, Abcam) was then used to incubate with the membranes. Signals were detected using ECL (Sigma-Aldrich, USA). Image J v1.48 software was used to normalize the expression levels of Bach-1 to GAPDH.

Statistical analysis

Mean values of 3 independent biological replicates were calculated. Comparisons between two groups were performed using paired t test. Comparisons among multiple groups were performed using one-way ANOVA and Tukey test. Correlation analysis was performed by linear regression. $P < 0.05$ was statistically significant.

Results

MIR17HG was downregulated in NSCLC

Differential expression of MIR17HG in NSCLC was first detected by exploring the TCGA dataset. It was observed that MIR17HG was downregulated in both adenocarcinoma (0.4 vs. 0.54) and squamous cell carcinoma (0.76 vs. 0.88), which are two major subtypes of NSCLC. To further confirm this, the expression levels of MIR17HG in both NSCLC and non-tumor tissues of the 60 NSCLC patients were measured by qPCR. Paired t test analysis showed that the expression levels of MIR17HG were significantly lower in NSCLC tissues than that in non-tumor tissues (Fig. 1, $p < 0.001$).

MiR-142-3p was positively correlated with MIR17HG

Expression of MiR-142-3p in both NSCLC and non-tumor tissues were also determined by qPCR. It was observed that the expression levels of miR-142-3p were also significantly

lower in NSCLC tissues than that in non-tumor tissues (Fig. 2 a, $p < 0.001$). Correlation analysis showed that the expression of miR-142-3p and MIR17HG were positively correlated across both NSCLC (Fig. 2 b) and non-tumor (Fig. 2 c) tissue samples.

Overexpression of MIR17HG decreased methylation of miR-142 and downregulated Bach-1

The effects of overexpressing MIR17HG on the methylation of miR-142 were assessed by MSP. Compared to cells transfected with pcDNA3.1 vector, cells transfected with MIR17HG expression vector showed obviously reduced methylation of miR-142 gene (Fig. 3 a). To further investigate the relationship between MIR17HG and miR-142-3p, cells were transfected with MIR17HG vector or miR-142-3p mimic. Overexpression of MIR17HG and miR-142-3p were confirmed by qPCR at 24 h post-transfection (Fig. 3 b, $p < 0.05$). It showed that overexpression of MIR17HG led to upregulation of miR-142-3p (Fig. 3 c, $p < 0.05$). Overexpression of MIR17HG led to downregulation of Bach-1, which is a target of miR-142-3p, at mRNA level (Fig. 3 e, $p < 0.05$). Representative western blot results were presented (Fig. 3 e, left, $p < 0.05$), and the results showed that overexpression of MIR17HG led to downregulation of Bach-1 at protein level as well (Fig. 3 e, right, $p < 0.05$).

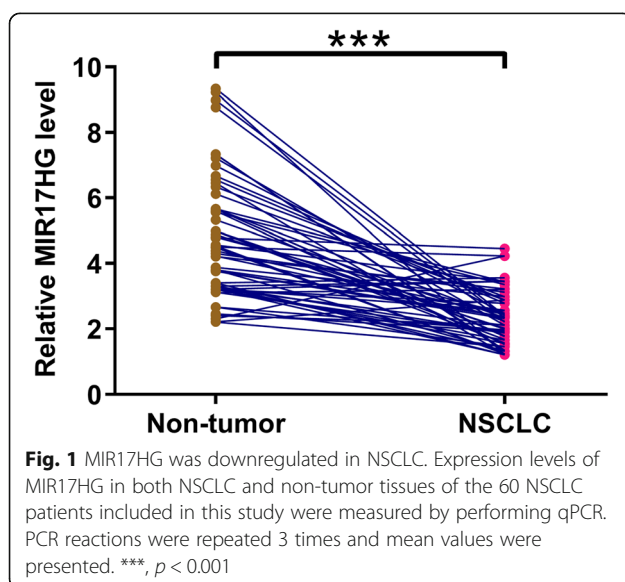
MIR17HG inhibited invasion and migration of cancer cells through miR-142-3p and Bach-1

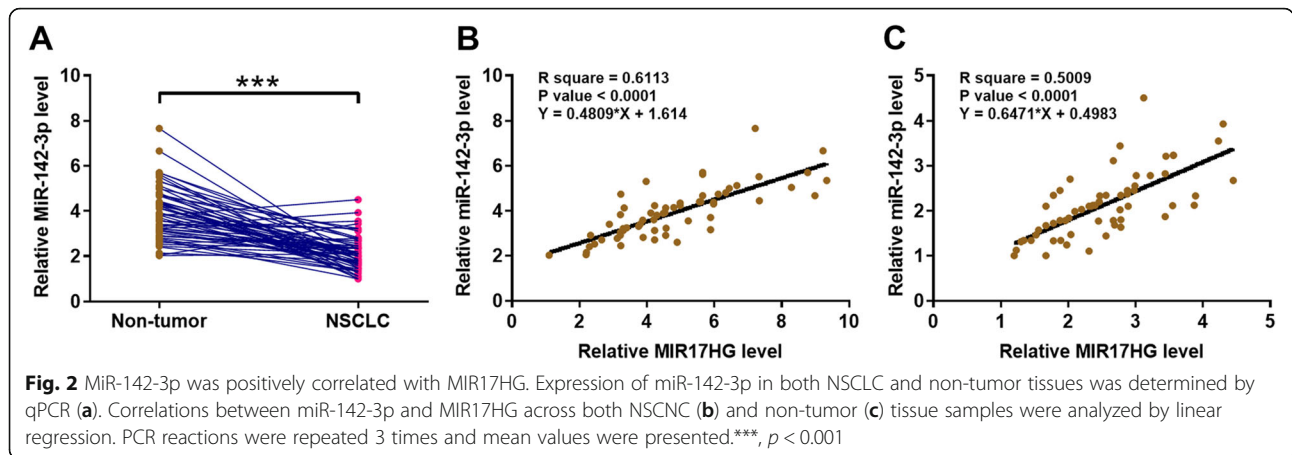
The effects of overexpressing MIR17HG, miR-142-3p and Bach-1 on the invasion (Fig. 4 a) and migration (Fig. 4 b) of NSCLC cells were assessed by Transwell invasion or migration assay. Compared to the Control (C) group or cells transfected with pcDNA3.1 vector or NC miRNA, overexpression of MIR17HG and miR-142-3p led to inhibited cancer cell invasion and migration. In contrast, overexpression of Bach-1 played an opposite role and attenuated the effects of overexpressing MIR17HG and miR-142-3p ($p < 0.05$). To further confirm the effects of MIR17HG, miR-142-3p and Bach-1 in regulating NSCLC cell invasion and migration, Transwell invasion (Supplemental Fig. 1A) and migration (Supplemental Fig. 1A) assays were repeated using another cell line H2126, and similar results were obtained.

Discussion

This study mainly investigated the roles of MIR17HG in NSCLC. We observed that MIR17HG was downregulated in NSCLC and may upregulate the expression miR-142-3p by reducing the methylation of this gene.

Expression patterns of MIR17HG in different types of cancer have not been well studied. It is reported that MIR17HG is the host of multiple oncogenic miRNAs that involved in the progression of cell death [12].

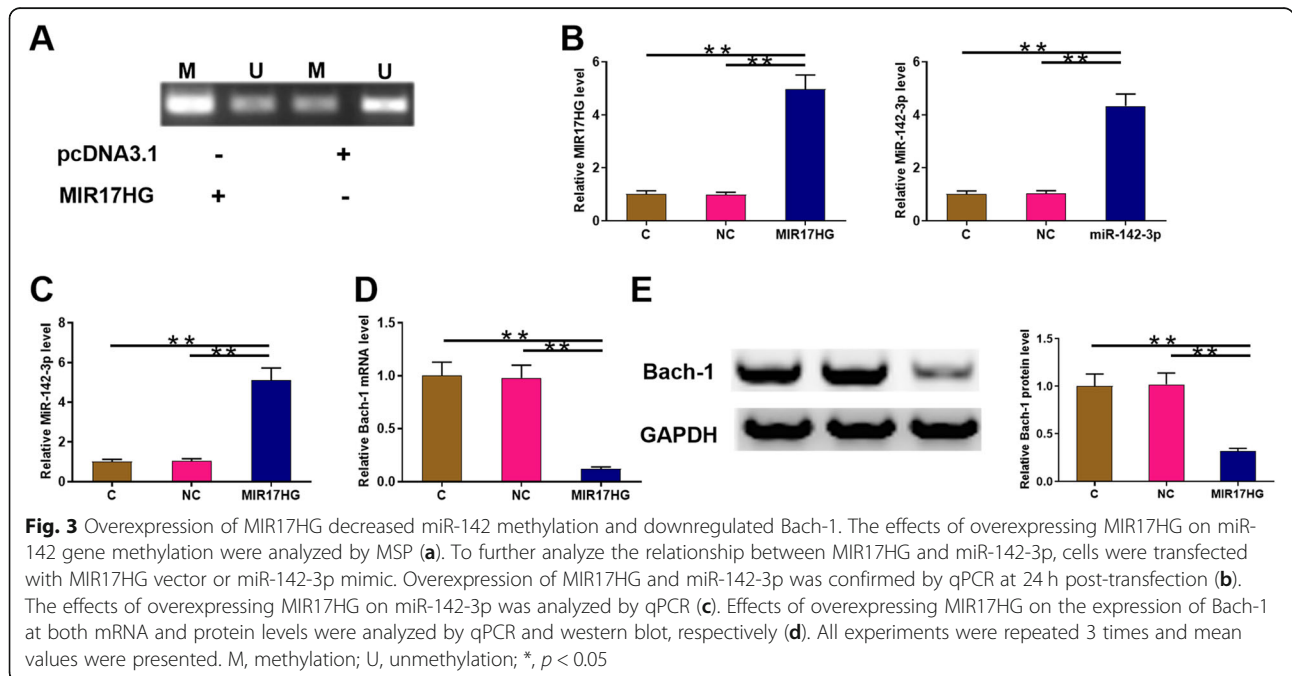




However, downregulation of MIR17HG was observed in many types of cancer after analyzing the TCGA dataset, such as acute myeloid leukemia (6.91 vs. 32.83), ovary cancer (0.88 vs. 1.77) and pancreas adenocarcinoma (0.84 vs. 1.64), which suggests the tumor suppressive roles of MIR17HG in these types of cancer. We also observed the slightly lower levels of MIR17HG in NSCLC tissues compared to that in non-tumor tissues by analyzing TCGA dataset. Significantly reduced expression levels of MIR17HG were observed in NSCLC tissues compared to that in non-tumor tissues in the 60 NSCLC patients included in this study. This is possibly due to the different populations included in these two datasets. Moreover, our study observed reduced invasion and

migration rates of NSCLC cells after the overexpression of MIR17HG. Therefore, MIR17HG may play tumor suppressive roles in NSCLC.

It is commonly observed that tumor suppressive miRNAs are frequently methylated in cancers [15–17]. For instance, miR-145 is regulated by DNA methylation in prostate cancer [15]. MicroRNA-34b/c is downregulated in gastric cancer through methylation-related pathways [16]. MiR-142-3p is a well-established tumor suppressive miRNA and has regulatory roles in multiple cell behaviors, such as migration and invasion. A recent study reported that miR-142 is hypermethylated in liver cancer [17]. Our study also observed the methylation of miR-142 in NSCLC cells. It is known that lncRNAs can regulate the methylation of miRNA genes [18].



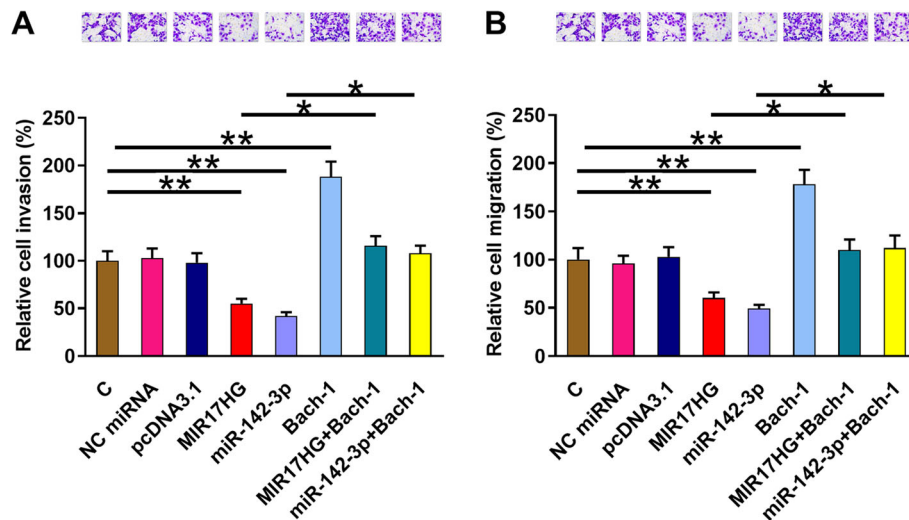


Fig. 4 MIR17HG inhibited invasion and migration of H2170 cells through miR-142-3p and Bach-1. The effects of overexpressing MIR17HG, miR-142-3p and Bach-1 on the invasion (a) and migration (b) of H2170 cells were assessed by Transwell invasion or migration assay. All experiments were repeated 3 times and mean values were presented. *, $p < 0.05$

In this study we observed that overexpression of MIR17HG reduced the methylation of miR-142-3p gene to upregulate its expression, and this expression regulation was involved in the invasion of migration of NSCLC cells. However, the mechanisms of methylation regulation of miR-142-3p by MIR17HG are still unclear.

It is known that Bact-1 in cancer biology is involved in the regulation of heme oxidation and oxidative stress. It is also linked to cancer invasion and metastasis [19]. Therefore, MIR17HG may directly regulate Bach-1 through miR-142-3p to regulate NSCLC cell invasion and migration. It is worth noting that we also tried to measure the expression levels of Bach-1 in paired tissues. However, Bach-1 is below detectable level in many non-tumor samples. This is possibly due to the low expression levels of Bach-1 in some patients. In our future studies, we will increase the sensitivity of qPCR to solve this problem.

Conclusion

In conclusion, MIR17HG was downregulated in NSCLC and may upregulate miR-142-3p through methylation pathway to promote its expression, thereby downregulating Bach-1 and suppress cancer cell invasion and migration.

Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s12890-020-1112-3>.

Additional file 1: Supplemental Fig. 1 MIR17HG inhibited invasion and migration of H2126 cells through miR-142-3p and Bach-1. The effects of overexpressing MIR17HG, miR-142-3p and Bach-1 on the invasion (A) and migration (B) of H2126 cells were assessed by Transwell invasion or migration assay. All experiments were repeated 3 times and mean values were presented. *, $p < 0.05$.

Abbreviations

ncRNAs: Non-coding RNAs; lncRNAs: Long (> 200 nt) non-coding RNAs; NSCLC: Non-small cell lung cancer

Acknowledgments

Not applicable.

Authors' contribution

SW and JHL designed and carried out the study. XL and XYL participated in experiments and statistical analysis. SW, JHL wrote the manuscript and revised the manuscript. All authors read and approved the final manuscript.

Funding

This study was supported by the grant from the Tianjin Medical University General Hospital Youth Incubation Fund (ZYYFY2017024).

Availability of data and materials

The analyzed data sets generated during the study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of General Hospital of Tianjin Medical University. The research has been carried out in accordance with the World Medical Association Declaration of Helsinki. All patients and healthy volunteers provided written informed consent prior to their inclusion within the study.

Consent for publication

All authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 29 August 2019 Accepted: 12 March 2020

Published online: 30 March 2020

References

- Mao Y, Yang D, He J, et al. Epidemiology of lung cancer. *Surg Oncol Clin N Am.* 2016;25(3):439–45.
- Bray F, Ferlay J, Soerjomataram I, et al. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 2018;68(6):394–424.

3. Islami F, Torre LA, Jemal A. Global trends of lung cancer mortality and smoking prevalence. *Transl Lung Cancer Res.* 2015;4(4):327–38.
4. Pirie K, Peto R, Green J, et al. Lung cancer in never smokers in the UK million women study. *Int J Cancer.* 2016;139(2):347–54.
5. Solomon BJ, Kim DW, Wu YL, et al. Final overall survival analysis from a study comparing first-line crizotinib versus chemotherapy in ALK-mutation-positive non-small-cell lung cancer. *J Clin Oncol.* 2018;36(22):2251–8.
6. Herbst RS, Morgensztern D, Boshoff C. The biology and management of non-small cell lung cancer. *Nature.* 2018;553(7689):446–54.
7. Steliga MA, Dresler CM. Epidemiology of lung cancer: smoking, secondhand smoke, and genetics. *Surg Oncol Clin N Am.* 2011;20(4):605–18.
8. Zhang Z, Lee JC, Lin L, et al. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nat Genet.* 2012;44(8):852–60.
9. Chan BA, Hughes BGM. Targeted therapy for non-small cell lung cancer: current standards and the promise of the future. *Transl Lung Cancer Res.* 2015;4(1):36–54.
10. Esteller M. Non-coding RNAs in human disease. *Nat Rev Genet.* 2011;12(12):861–74.
11. Lopez MF, Niu P, Wang L, et al. Opposing activities of oncogenic MIR17HG and tumor suppressive MIR100HG clusters and their gene targets regulate replicative senescence in human adult stem cells. *NPJ.* 2017;3(1):7.
12. Mansoori B, Mohammadi A, Ghasabi M, et al. miR-142-3p as tumor suppressor miRNA in the regulation of tumorigenicity, invasion and migration of human breast cancer by targeting Bach-1 expression. *J Cell Physiol.* 2019;234(6):9816–25.
13. Mao JT, Xue B, Smoake J, et al. MicroRNA-19a/b mediates grape seed procyanidin extract-induced anti-neoplastic effects against lung cancer. *J Nutr Biochem.* 2016;34:118–25.
14. Xu J, Meng Q, Li X, et al. Long noncoding RNA MIR17HG promotes colorectal Cancer progression via miR-17-5p. *Cancer Res.* 2019;79(19):4882–95.
15. Suh SO, Chen Y, Zaman MS, et al. MicroRNA-145 is regulated by DNA methylation and p53 gene mutation in prostate cancer. *Carcinogenesis.* 2011;32(5):772–8.
16. Suzuki H, Yamamoto E, Nojima M, et al. Methylation-associated silencing of microRNA-34b/c in gastric cancer and its involvement in an epigenetic field defect. *Carcinogenesis.* 2010;31(12):2066–73.
17. Yu Q, Xiang L, Yin L, et al. Loss-of-function of miR-142 by hypermethylation promotes TGF- β -mediated tumour growth and metastasis in hepatocellular carcinoma. *Cell Prolif.* 2017;50(6):e12384.
18. Liu H, Fang L, Cheng Y, et al. LncRNA PVT1 regulates prostate cancer cell growth by inducing the methylation of miR-146a. *Cancer Med.* 2016;5(12):3512–9.
19. Davudian S, Mansoori B, Shajari N, et al. BACH1, the master regulator gene: a novel candidate target for cancer therapy. *Gene.* 2016;588(1):30–7.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

